# Phylogeography of the European rabbit (*Oryctolagus cuniculus*) in the Iberian Peninsula inferred from RFLP analysis of the cytochrome *b* gene

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We studied mitochondrial DNA variation in the European rabbit through the examination of restriction fragment length polymorphism in 526 individuals from 20 locations spread across the Iberian Peninsula. Digestion with eight enzymes of a 1120-bp fragment comprising most of the cytochrome *b* gene resolved 38 different haplotypes. These haplotypes were distributed in two highly divergent clades, with different but overlapping geographical distributions, and with comparable levels of within-clade variation. The overall phylogeographical pattern suggests a history of long-term regional isolation of two groups of rabbit populations, compatible with the recognition of two subspecies within the Iberian Peninsula, followed by recent contact and admixture. The underlying cause is sought in the alternation of glacial and interglacial periods in the late Pleistocene.

**Keywords:** cytochrome b, European rabbit, glaciations, Iberian Peninsula, Oryctolagus cuniculus, phylogeography.

#### Introduction

Both wild and domestic European rabbit belong to the single species Oryctolagus cuniculus, which is the sole extant representative of the genus *Orvetolagus*. Palaeontological data indicate limited morphological diversification of the genus since its appearance at the end of the Miocene (6.5 MYA) (Lopez-Martinez, 1989). Fossil remains from the middle Pliocene led to the recognition of two species, O. lacosti in southern France and northwestern Italy and O. laynensis in the Iberian Peninsula. This latter form is thought to be the origin of the extant O. cuniculus (Lopez-Martinez, 1989). Archaeozoological data indicate that the original range of the rabbit was restricted to the Iberian Peninsula and the south of France up to river Loire (Callou, 1995). During the Middle Ages, humans started to have a major impact on rabbit distribution by transporting animals to many other geographical areas and rabbits can nowadays be found almost all over the world (Flux, 1994). Two subspecies are recognized, *Oryctolagus cuniculus cuniculus* which is present in France and in most areas where they were introduced by humans and *O. c. algirus* that occupies the Iberian Peninsula, North Africa, Mediterranean and Portuguese Atlantic islands (cf. Ellerman & Morrison-Scott, 1951; Saint-Girons, 1973; Wilson & Reeder, 1992).

The study of restriction fragment length polymorphism (RFLP) of the entire rabbit mtDNA molecule revealed the existence of two very distinct maternal lineages (Ennafaa et al., 1987; Biju-Duval et al., 1991). Lineage A was found in the south-west of the Iberian Peninsula and lineage B in the rest of Europe (including northern Spain) and also in domestic breeds. Further work revealed a total of 22 haplotypes in 12 wild populations from the Iberian Peninsula, southern France and the Mediterranean island of Zembra (Tunisia), and one French domestic stock (Monnerot et al., 1994). A clear subdivision into two groups was also found when nuclear genes were studied (van der Loo et al., 1991, 1999; Ferrand, 1995), with a geographical distribution that largely coincides with that of the two mitochondrial clades A and B. The geographical partitioning of these two groups can be superimposed, with few exceptions, on the distribution of the two

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subspecies based on classical taxonomy. Thus clade A was tentatively associated with *O. c. algirus* and clade B with *O. c. cuniculus*, as well as the corresponding nuclear gene pools (Biju-Duval *et al.*, 1991; van der Loo *et al.*, 1991; Ferrand, 1995; Branco & Ferrand, 1998).

Although the natural range distribution of the rabbit once comprised southern France up to river Loire (Callou, 1995), these populations are not relevant for the understanding of the early steps of rabbit history. As shown by ancient mtDNA, these populations were already less variable than their Iberian counterparts and human intervention has resulted in noticeable changes of their mtDNA genetic composition (Hardy et al., 1995; Loreille et al., 1997). Therefore, French populations (or any others to the north of the Pyrenees) tell us, mostly, the recent history of human-mediated colonization and expansion that occurred from the Middle Ages to the present. Thus, the major clues for the evolutionary history of the rabbit have to be looked for in its very original range, i.e. the Iberian Peninsula, by performing a detailed analysis of rabbit genetic structure and diversity in this area, not yet extensively sampled in previous studies.

In this work, a comprehensive survey of rabbit mtDNA diversity in the Iberian Peninsula by means of RFLP analysis will be presented. The strong geographical partitioning of mtDNA clades A and B is confirmed by our data, and the occurrence of both maternal lines in several populations is also described. This phylogeographical pattern suggests that two groups of popula-

tions were isolated for a certain time and later expanded and overlapped. The possibility that this separation resulted from retreat into glacial refugia during Quaternary ice-ages and that the present mixed populations resulted from secondary contact due to postglacial expansion and recolonization will be discussed. Finally, these two distinct groups will be related with the subspecies *algirus* and *cuniculus* and, subsequently, a re-evaluated geographical distribution in the Iberian Peninsula will be suggested.

#### Materials and methods

#### Sample collection

A total of 526 wild rabbits were sampled from 20 locations across the Iberian Peninsula (Fig. 1, sampling sites are named after the closest city or town except for Cabreira, Infantado, Doñana National Park and Las Lomas). Outside the hunting season rabbits were caught by ferreting and blood was collected from the marginal ear vein in EDTA-coated tubes. During the hunting season a portion of liver was taken from freshly killed animals. Samples were stored at -20°C until use.

#### DNA extraction

Total DNA was obtained from liver or blood by proteinase K digestion, phenol/chloroform extraction and ethanol precipitation using classical techniques. In

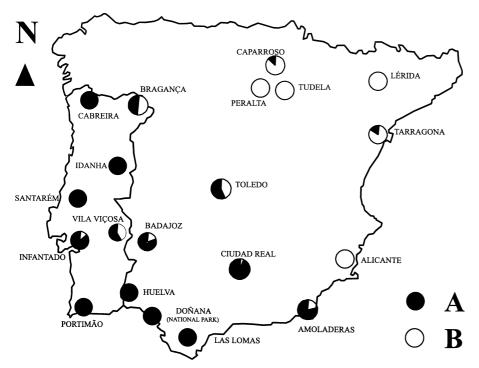


Fig. 1 Geographical distribution of lineages A and B in the Iberian Peninsula. Locations are named after the town or city closest to the sampling site (except for Cabreira, Doñana, Infantado and Las Lomas).

samples from Las Lomas and Badajoz, mtDNA was obtained from liver purified on CsCl gradients (Ennafaa et al., 1987; Biju-Duval et al., 1991).

#### **Amplification**

A 1120-bp sized fragment of mtDNA, including most of the cytochrome b gene, was amplified with Tag Goldstar according to instructions of the manufacturer (Eurogentec). The reaction mixture consisted of 50 µL Eurogentec buffer, 150  $\mu$ M dNTPs and 1  $\mu$ M primers. Following 3 min of denaturation at 96°C, PCR was done in a Kontron cycler over 40 cycles of 1 min at 94°C, 1 min at 55°C and 1 min at 72°C. After the last cycle PCR was incubated a further 3 min at 72°C. The primers used were cytb8 (CCCCA TCCAA CATCT CTGCT TGATG AAA; 14242-14269) (Barome et al., 1998) and th3 (GTCTT GTAAG CCAGG AATGG AG; 15340-15361) (Hardy et al., 1994). The numbers indicate the nucleotide positions at the 5' and 3' ends, respectively, relative to the rabbit mitochondrial genome (accession number AJ001588, Gissi et al., 1998).

#### DNA sequencing

The sequence of the 1120-bp fragment for an individual carrying haplotype A2Rba was obtained by direct sequencing using Sequenase PCR Products sequencing kit (USB). This sequence was used as reference in restriction site mapping of lineage A (accession number AJ243096).

#### Restriction site analysis

Amplified mtDNA was digested with eight restriction enzymes (AluI, BsaI, DdeI, FokI, HaeIII, MseI, TaqI and Tsp509I) following instructions provided by the suppliers. The fragments obtained were separated by electrophoresis on 3% Nusieve agarose gels. The gels were immersed in an ethidium bromide solution 0.5% (w/v) for 10 min and RFLP patterns were visualized under UV light. Profiles are given in Table 1 with all generated fragments ordered by their relative position along the sequence.

## Haplotype nomenclature

For simplicity, the 38 observed haplotypes will be referred to by the number given in the first column of Table 2. Sequences corresponding to haplotypes 1 and 19 (accession numbers AJ243197 and AJ243096) were used as references in the restriction site mapping of lineage B and lineage A, respectively. Distinct single endonuclease patterns were identified by a specific letter

in order of appearance. Each individual was assigned a multiletter code describing its haplotype for the eight endonucleases (Table 2). Haplotypes described earlier (Ennafaa et al., 1987; Biju-Duval et al., 1991; Monnerot et al., 1994) were retyped and allocated to 12 haplotypes as shown in Table 2.

## Data analysis

The MAPSORT subroutine of GCG (Devereux, 1989) was used to locate the site gain or loss for each new endonuclease profile. Haplotype and nucleotide diversity and nucleotide divergence were calculated using the REAP package (McElroy et al., 1991). Maximum parsimony was calculated using the heuristic search option of the PAUP software package (Swofford, 1998). Statistical parsimony was used to choose the best network among 100 networks of equal length. The 'show patristic distances' option of PAUP was used to identify the maximum parsimony network closest to the real data (see Templeton et al., 1987, 1992; Templeton & Sing, 1993, for details).

#### Results

Five hundred and twenty-six wild rabbits from 20 Iberian locations were screened with eight restriction enzymes generating 52 restriction sites which discriminated 38 different haplotypes, of which 26 were new. The 38 haplotypes clustered into the two major lineages A and B, which are concordant with the two main groups previously described (Biju-Duval et al., 1991; Monnerot et al., 1994). Twenty-one haplotypes were of type A and 17 of type B. The phylogenetic relationships were represented by the maximum parsimony network depicted in Fig. 2(a) and this does not differ importantly from that obtained using the distances approach (genetic distances were computed according to Nei & Tajima, 1981; results not shown). Clades A and B were separated by at least 14 mutations (nucleotide divergence 11.9%) and they exhibited similar levels of withinclade polymorphism with nucleotide diversities of 1.1% and 1.4%, respectively. The number of different haplotypes per population averaged 4.6 (range 1–7). Across individual rabbits, 329 carried type A mtDNA and 197 carried type B.

#### Geographical distribution of clades

Globally, the two mtDNA clades A and B show a highly structured geographical pattern of distribution (Fig. 1). Clade A was fixed or almost fixed in all south-western locations whereas clade B was almost the only one present in the north and east. Both clades were

**Table 1** Patterns obtained by cleavage with eight endonucleases of an amplified fragment (1120 bp) of rabbit mtDNA comprising most of cytochrome b. The fragments generated by eight restriction enzymes are ordered by their position along the sequence

Endonuclease					Profiles				
AluI	A 810 310	B 504 289 17 310	C 504 306 310	D 504 616	E 414 90 616	F 1120	G 504 333 283	H 87 417 306 310	
BsaI	A 258 421 441	B 679 441	C 258 862	D 1120	E 258 421 399 42				
DdeI	A 83 327 15 150 333 212	B 83 327 15 81 69 333 212	C 83 327 15 150 45 36 464	D 410 15 81 69 45 36 464	E 83 327 15 150 45 36 252 212	F 83 342 150 45 36 464	G 83 327 15 81 69 45 36 464		
FokI	A 270 540 18 174 118	B 179 91 540 18 174 118	C 153 315 112 248 152 140	D 153 315 112 230 18 152 140	E 153 315 112 400 140	F 270 333 153 54 18 174 118	G 270 354 186 18 174	H 270 333 207 18 174 118	I 153 26 91 540 18 174 118
HaeIII	A 46 30 153 723 168	B 46 183 723 168	C 46 183 891	D 46 1074	E 46 183 221 670				
MseI	A 151 362 205 402	B 513 607	C 513 205 402	D 513 97 108 402	E 718 402	F 513 205 122 280			
TaqI	A 1048 72	B 442 606 72	C 415 633 72						
<i>Tsp</i> 509I	A 58 6 450 48 558	B 58 6 450 606	C 58 6 88 362 606	D 58 6 303 147 606	E 58 6 88 362 93 513	G 58 6 88 362 69 537			

1													Locations	ons				1					
PREMETER	No.	Reference (*)	Composite	BRA	CAB		INF	SAN	VV													PRT	Total
Berbait Bish bish Andama, and a control of the cont	1	B1Rba (B1)	AAAAAAA								0.09	0.27		70.0		.12							13
BKBNb (Ne Br) (MANANA)         0.90         0.30         0.13         0.15         0.1	7	B3Rba (B3 B4)	BAAAAAAA				0.13		0.40		98.0					.24	0.1	6					57
BRISH GRAMARA GILLAMANARA GILL	3	B6Rba (B6 B7)	CAABAAAA	0.08									90.0			.33							18
BIRDIA GRIJAN AARAAAAA AABAAAAA AABAAAAA AABAAAAA AABAAAAA	4	B8Rba (B8 B9 B10)	AABAAABB	0.11						0.50		0.53		.72									43
BRN1         AMBAAANAA         6.20         6.20         6.07         6.02         6.03         6.03         6.03	5	B11Rba (B11)	CBAAAABA									0.07	_		.13								6
BRB2         AAABAAAAB         0.03         A.B. AAABAAAB         0.07         0.03         A.B. AAABAAAB         0.05         A.B. AAABAAAB         A.B. AAABAAAB         0.05         A.B. AAABAAAB         A.B. AAABAAAB         A.B. AABAAAB         A.B. AABAAAB <t< td=""><td>9</td><td>BRb1</td><td>AABAAABA</td><td></td><td></td><td></td><td></td><td></td><td></td><td>0.20</td><td></td><td></td><td></td><td>J</td><td></td><td>70.0</td><td></td><td>0.0</td><td>5</td><td></td><td></td><td></td><td>12</td></t<>	9	BRb1	AABAAABA							0.20				J		70.0		0.0	5				12
BRB   Abraham	7	BRb2	НААНАААА									0.07		.03									7
BRB-b  CDAAAAAA   25	∞	BRb3	AABAAAAB	0.03																			-
BRISH         ACMAAABA         25         4 CMAAABA         1 CMAABAA         1 CMAAABA         1 CMAAABA         1 CMAAABA         1 CMABAAAA         1 CMABAAAA         1 CMABAAAA         1 CMAAAAAA         1 CMAAAAAA         1 CMAAAAAA         1 CMAAAAAA         1 CMAAAAAA         1 CMABAAAAAAAAAAA         1 CMABAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA	6	BRb4	CDAAAABA											Ū	.05								2
BR bb	10	BRb5	ACBAAABA	0.26																			10
BRb1         CAAFAAAA         0.05         0.07         0.10         A. A	Ξ	BRb6	CAABAACA							0.05													-
BRbb         AAAGAABA         007         010         A.BGABA         A.BGAABAA         A.BGAABAA         A.BGAABAAA         A.BGAABAAA         A.BGAABAAAA         A.BBB BB	12	BRb7	CBAFAAAA							0.05													_
BRb10         CAAAAAAAAA         0.03         6.10         1.10	13	BRb8	AABGAABA									0.07											_
BRb11         CEATAAAA         0.10         0.24         0.10         0.24         0.10         0.24         0.10         0.24         0.25         0.29         0.23         0.13         0.24         0.25         0.20         0.23         0.13	14	BRb10	CAAAAAA								0.05		92.(			0.]	0						15
BRb12         BBAAAAAA         0.10         0.10         0.44         0.24         0.23         0.23         0.23         0.13	15	BRb11	CEAIAAAA													0.]	0.						-
BRb13         CAABAABA         0.03         0.10         0.44         0.25         0.26         0.27         0.29         0.10         0.10         0.44         0.25         0.20         0.27         0.25         0.27         0.29         0.14         0.00         0.05         0.05         0.04         0.00         0.05         0.05         0.04         0.00         0.05         0.05         0.04         0.00         0.05	16	BRb12	BBAAAAAA													.24							10
AlRba (Al A S AT)         DBCCCBBBC	17	BRb13	CAABAABA	0.03																			_
ARRba (A2 A4)         DBCCCCCCC         0.04         0.05         0.05         0.05         0.05         0.05         0.05         0.05         0.05         0.05         0.05         0.05         0.05         0.05         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.13         0.05         0.13         0.05         0.13 <td>18</td> <td>A1Rba (A1 A5 A7)</td> <td>DBCCBBBC</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>0.10</td> <td></td> <td></td> <td></td> <td>Ū</td> <td>4.</td> <td></td> <td>0.2</td> <td></td> <td></td> <td></td> <td>0.33</td> <td>0.18</td> <td>99</td>	18	A1Rba (A1 A5 A7)	DBCCBBBC							0.10				Ū	4.		0.2				0.33	0.18	99
A3Kba (A3)         DBCDBBD         A3Kba (A3)         DBCCDBBC         A3Kba (A8)         BCCBBC         A3Kba (A8)         A3Kba (A8)         BCCCBBC         A3Kba (A8)         A3Kba (A9)         BCCCBBC         A3Kba (A9)         BCCCBBC         A3Kba (A9)         BCCCBBC         A3Kba (A9)	19	A2Rba (A2 A4)	DBCCCCBC	0.29	0.48		0.29	0.04	90.0	0.05				•	.05		0.0	9		0.27	0.26		27
ARRba (Ab)         EBCCBBBC           ASR Ra (As)         DBCCCCBC         0.04         0.05         0.05         0.05         0.13         0.13           AINBba (AlI)         DBCCCCBC         0.04         0.06         0.06         0.10         0.35         0.12         0.10         0.12         0.10         0.10         0.11         0.12         0.10         0.12         0.12         0.10         0.12         0.12         0.10         0.12         0.13         0.13         0.13         0.13         0.13         0.12         0.12         0.10         0.12         0.13	20	A3Rba (A3)	DBCDBBBD																		0.15		∞
A8Rba (A8)         DBCCDCBC         4006	21	A6Rba (A6)	EBCCBBBC																		0.13		7
A9Rba (A9)         FBDCCCBC         0.06         0.06         0.05         0.05         0.12         0.12         0.13         0.12         0.13         0.14	22	A8Rba (A8)	DBCCDCBC																		0.13		7
AIIRba (AII)         GBCCBBC         0.04         0.06         0.05         0.10         0.15	23	A9Rba (A9)	FBDCCCBC														0.0	9					-
AIIRbat (AII)         DBECCCBC         0.04         0.06         0.80         0.19         0.05         0.12         0.12         0.12         0.11         0.12         0.12         0.12         0.13         0.14         0.14         0.15         0.14         0.15         0.14         0.18 <td>24</td> <td>A10Rba (A10)</td> <td>GBCCBBBC</td> <td></td> <td>4</td>	24	A10Rba (A10)	GBCCBBBC																				4
ARbJ         DBECBBBC         0.09         0.10         0.12         0.10         0.25         0.69         0.41           ARbJ         DBCCCBB         0.21         0.23         0.83         0.46         0.12         0.10         0.06         0.06           ARbJ         DBCCCBB         0.21         0.21         0.21         0.21         0.04         0.06         0.06         0.06         0.06         0.06         0.06         0.06         0.06         0.06         0.04         0.06         0.04         0.04         0.08         0.06         0.08         0.06         0.08         0.06         0.08         0.08         0.06         0.08         0.08         0.06         0.08         0.08         0.06         0.08         0.08         0.08         0.08         0.08         0.08         0.08         0.08         0.08         0.08         0.08         0.08         0.08         0.08         0.09	25	AllRba (All)	DBECCCBC				0.04		0				90.0			3.0			ς.	0.58		0.12	31
ARB2         DBCCCBBB         0.10         0.12         0.10         0.12         0.41           ARB4         DBCCCCBB         0.21         0.52         1.00         0.33         0.83         0.46         0.10         0.08         0.06           ARB5         DBCCCBBB         0.21         0.13         0.83         0.83         0.83         0.83         0.04         0.06           ARB6         DBCCCBB         0.21         0.21         0.21         0.04         0.04         0.08           ARB1         DBCCCBC         0.04         0.08         0.04         0.04         0.04         0.04           ARB1         DBCCCBB         0.04         0.04         0.04         0.04         0.04           ARB1         DBCCCBB         0.04         0.04         0.04         0.04         0.04           ARB1         DBCCCBB         0.04         0.04         0.04         0.04         0.04	26	ARbl	DBFCCCBC						0.00					,	•								. n
ARb3         DBCCCBB         0.21         0.25         0.09         0.41           ARb4         DBCCCCBB         0.21         0.23         0.46         0.10         0.06           ARb5         DBCCCCBB         0.13         0.13         0.13         0.04         0.04           ARb9         FBCCCBB         0.21         0.21         0.04         0.04         0.04           ARb1         DBCCCBCB         0.04         0.08         0.04         0.08         0.04           ARb1         DBCCCBBC         0.04         0.04         0.04         0.04         0.04	/7 6	AK62	DBECBBBC										5		01.			•				-	4 5
ARb4         DBCCCBB         0.13         0.13         0.00	87 6	ARb3	DBCCCCBE	5	63	5	,,	000	77			-	7.17					7.0				0.41	75 6
ARD         DBCCEBB         0.13         0.10         0.08           ARb         DBCCEBB         0.21         0.25         0.04           ARb         FBCCCBB         0.01         0.04         0.08           ARb         DBCCCEBC         0.08         0.08           ARb         DBCCCBB         0.04         0.06           ARb         DBCCCBB         0.04         0.06           ARb         ARb         0.06         0.08           ARb         DBCCCBB         0.04         0.06	67 6	AND4	DECCCEBB	17.0	0.32	1.00	CC.0	0.03	0.40									Ċ				0.00	ţ -
ARB         DBCCECBB         0.25           ARb         DBCCECBB         0.21         0.04           ARb         DBCCCEBC         0.08         0.08           ARb         DBCCCBBC         0.08         0.04           ARb         DBCCCBBC         0.04         0.06           ARb         DBCCCBBC         0.04         0.06           ARb         DBCCCBBC         0.06         0.08	£ 5	AK65	DBCEBBBC					,										0.1					4 r
ARbb         FBCCCBB         0.21         0.23           ARb10         EBCCBBG         0.04         0.08           ARb11         DBCCCEBC         0.08         0.04           ARb12         DBCCCBB         0.04         0.06           ARb13         DBCCCBBC         0.06         0.18           ARb14         DBCECFBE         0.18	15	ARb/	DECCLEBB					0.13										·	4				n 4
ARbit         EBCCBBG         0.04           ARbit         DBCCCEBC         0.08           ARbit         DBCCCBB         0.04           ARbit         DBCCCBB         0.04           ARbit         DBCCCBB         0.06           ARbit         DBCCCBB         0.06           ARbit         DBCCCFBE         0.18	ر در	ARDS APL0	DECCEUBB				5											7.0	0				n u
ARbit         EBCCBBG         0.04           ARbit         DBCCCEBC         0.08           ARbit         DBCCCBB         0.04           ARbit         DBCCCBB         0.06           ARbit         DBCCCFBE         0.18	6.	AKD9	FBCCCCBB				0.71													0			o .
ARbit         DBCCCEBC         0.08           ARbit         DBCCBCBB         0.04           ARbit         DBCCCBBC         0.06           ARbit         DBCCCFBE         0.18	45	AKblu	EBCCBBBG																	0.04			<u> </u>
ARb12         DBCCBCBB         0.04           ARb13         DBCCCBBC         0.06           ARb14         DBCECFBE         0.18	35	ARb11	DBCCCEBC																	0.08			7
ARb13 DBCCCBBC 0.06 ARb14 DBCECFBE 0.18	36	ARb12	DBCCBCBB																	0.04			-
ARb14 DBCECFBE 0.18	37	ARb13	DBCCCBBC																			90.0	-
	38	ARb14	DBCECFBE																			0.18	n

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 Table 2
 (Continued)

												Locations	ions										
No.	No. Reference (*) Composite BRA CAB	Composite	BRA	CAB	ID	INF	SAN	W	CAP	LER	PER	TAR	TUD	TOL	ALI	ID INF SAN VV CAP LER PER TAR TUD TOL ALI AMO BA CRE DON HUE LL PRT	BA	CRE I	NOC	HUE	TT	PRT T	Total
Haploty	Haplotype diversity		0.805 0.522	0.522	0.000	0.775	0.312	0.639	0.726	0.255	9.676	0.419	0.47	0.771	0.775	0.000 0.775 0.312 0.639 0.726 0.255 0.676 0.419 0.47 0.771 0.775 0.378 0.85 0.811 0.48	.85 (	0.811 0		0.609	0.781 (	0.794	
SE			0.0307	0.0307 0.0327		0.0423	0.1146	0.047	0.0917	0.1161	0.1049	0.1413	0.1083	0.0547	0.0289	0.00000 0.0423 0.1146 0.047 0.0917 0.1161 0.1049 0.1413 0.1083 0.0547 0.0289 0.1813 0.0469 0.0467 0.094 0.0821 0.0257	.0469 (	0.0467	.094	0.0821	.0257 (	0.0748	
Nucleot	Vucleotide diversity (%)		7.536 0.273	0.273	0.000	3.406	0.000 3.406 0.175 6.556 4.26	6.556	4.26	0.199	1.121	4.15	0.949	899.9	0.942	0.199 1.121 4.15 0.949 6.668 0.942 4.486 5.149 2.555 0.772	5.149	2.555 0	.772 0	0.57	1.151	1.119	
N			38	38 23	30	24	23	35	20	20 22 15 17 29	15	17	29	39	42	39 42 10 16 20	16	20	26	26 26 54 17	54		526

BRA, Bragança; CAB, Cabreira; ID, Idanha; INF, Infantado; SAN, Santarém; VV, Vila Vicosa; CAP, Caparroso; LER, Lérida; PER, Peralta; TAR, Tarragona; TUD, Tudela; TOL, Toledo; ALL, Alicante; AMO, Amoladeras; BA, Badaioz; CRE, Ciudad Real; DON, Doñana; HUE, Huelva; LL, Las Lomas and PRT, Portimão In the text haplotypes are referred to by the numbers given in the first column.

genome haplotypes from Biju-Duval et al. (1991) are given in parentheses (\*) in column 'Reference'

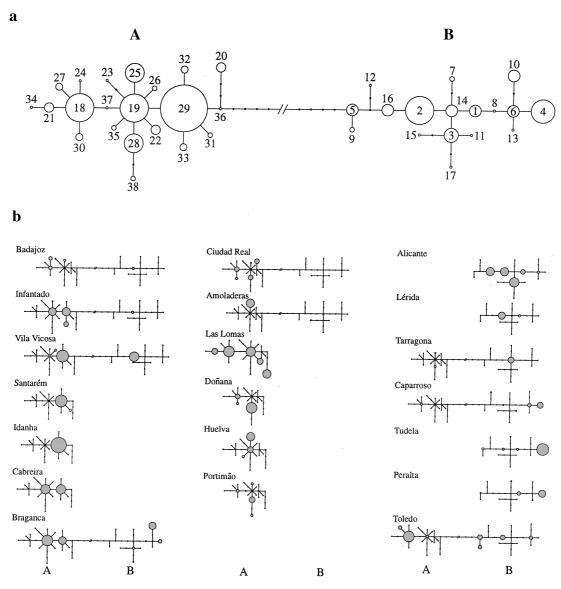
Haplotypes 1 to 17 were from lineage B; haplotypes 18 to 38 were from lineage A.

approximately equally common in three more or less centrally located populations (Toledo, Bragança and Vila Vicosa).

The high estimated average of nucleotide divergence (4.4%) obtained among populations reflects the level of genetic structuring and, mainly, the coexistence in several populations of haplotypes from clades A and B. Maximum nucleotide diversity ranged from 1.2% in Las Lomas and 1.1% in Peralta for clade A and clade B, respectively, to 7.5% for the mixed population of Bragança (Table 2).

# Patterns of haplotype distribution and population diversity

Six out of the 38 haplotypes have a wide geographical distribution. On the other hand, 24 are present only in a single location, of which 10 are in a single individual. These private haplotypes were distributed among 14 of the 20 locations surveyed. The distribution of the 38 haplotypes over the 20 locations is summarized in Fig. 2(b). The observed mtDNA diversity is superimposed over the maximum parsimony network of Fig. 2(a) representing only the haplotypes detected in each location. The observation of Fig. 2(b) reveals a clear trend in the interior vs. exterior position of haplotypes in the network. For clade B, the trend is from coastal Mediterranean locations (Alicante, Tarragona, Lérida) with interior positioned haplotypes, inland to the Ebro valley (Caparroso, Tudela, Peralta), and further westwards to north-eastern Portugal (Bragança) with exterior haplotypes. A similar pattern though less conspicuous, appears to be present for clade A from Las Lomas with interior haplotypes, northwards in the direction of Badajoz and Ciudad Real with haplotypes increasingly positioned towards the exterior. Southern locations proved to be the most variable and are characterized by the widespread occurrence of haplotypes 18, 19 and 25. Lower levels of variation were found along the western coast in a northward direction with one location monomorphic for haplotype 29 which is restricted to this region (Table 2). Inspection of Fig. 2(b) reveals that this particular haplotype characterizes all western locations, and that two of their few private haplotypes are derived from it. The highest level of variability within clade B was observed in the south-eastern coast and in northern Spain. The location of Alicante is characterized by a balanced distribution of five different haplotypes, whereas haplotype 4 dominates the Navarra locations (Caparroso, Peralta and Tudela). The lowest levels of diversity were found in the northeastern coast, in Lérida and Tarragona, where haplotypes 2 and 14, respectively, predominate. Clades A and B were found to coexist in nine locations, but only in



**Fig. 2** (a) Maximum parsimony network representing the phylogenetic relationships between the 38 mtDNA types. Circle size is proportional to the number of individuals in the total sample. Black points represent potential intermediates. (b) Parsimony networks for populations. Detected haplotypes are represented by grey circles with size proportional to the number of individuals (refer to the previous figure for haplotype number). Unobserved haplotypes and potential intermediates are represented by black points.

three of them were they equally abundant (Bragança, Toledo and Vila Viçosa). The six remaining locations have either clade A or clade B predominating (Table 2).

#### **Discussion**

#### Evidence for only two distinct mtDNA lineages

The fact that all haplotypes found in the original range of the rabbit species cluster within only two clades is a strong evidence that there are no more than two divergent lineages within *O. cuniculus*. Thus, our data confirm and further extend previous work in which a strong geographical subdivision of the two clades as well as the occurrence of admixed populations were described (Biju-Duval *et al.*, 1991; Monnerot *et al.*, 1994). The detection of both lineages A and B in Badajoz led Monnerot *et al.* (1994) to suggest the possibility of a contact zone, without, however, excluding the possibility that mixed populations resulted from introductions.

The detailed analysis of the distribution of the two distinct mtDNA clades throughout the Iberian

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Peninsula revealed a clear geographical pattern whereby south-western populations are fixed or nearly fixed for one lineage and north-eastern populations for the other. Rabbits of both mtDNA clades coexist in several locations in a geographically intermediate region (Fig. 1). The deep genetic differentiation observed, together with the contiguous and marginally overlapping ranges can be explained in a straightforward way by the allopatric origin of two monophyletic groups (Avise, 1989). Indeed, the observed haplotype distribution is in line with the persistence of rabbit populations in two main refugial areas during Pleistocene glaciations, followed by postglacial expansion after the last glacial maximum approximately 20 000 years ago. An alternative hypothesis of single origin would involve a highly biased process of lineage sorting and extinction, which in the present case seems to be much less probable.

Several phylogeographical studies provide evidence for the existence of glacial refugia in the Iberian Peninsula during the Quaternary (for example Cooper et al., 1995; Hewitt, 1996; Dumolin-Lapègue et al., 1997; Comes & Abbot, 1998; Taberlet et al., 1998; Sinclair et al., 1999). Yet, a remarkable feature of the two mtDNA clades observed in rabbits is their deep divergence, suggesting a long-time separation. Using an average rate of mammalian mtDNA divergence, Biju-Duval et al. (1991) estimated a divergence time of 2 MY for clades A and B. This places their origin on a Pliocene/Pleistocene boundary dated by several authors at 1.6–2.0 MYA (Williams et al., 1993), predating the more recent glaciation events.

Our RFLP analysis of cytochrome b yielded an estimated nucleotide divergence of 11.9% between clades A and B, which is in contrast with a previously reported value (4.2%) obtained from the entire mtDNA molecule (Biju-Duval et al., 1991). This may be due to the use of different nucleotide sampling regimes, and accepting the published calibration (Biju-Duval et al., 1991) we calculate a divergence rate of 6% per MY from our data. Within clades, nucleotide diversities (1.1% within clade A and 1.4% within clade B) would then correspond to a most recent common mitochondrial ancestor (MRCMA) at about 183 000 and 233 000 years, for clade A and clade B, respectively. Despite the technical differences, a very similar estimate for MRCMA was made by Biju-Duval et al. (1991) based on the analysis of the southern Spanish population of Las Lomas.

#### Haplotype phylogeny and population history

The extensive nucleotide diversity observed in both clades in which a one-step evolutionary branching

pattern is predominant suggests that a common historical factor – perhaps related with a climatic amelioration and a significant improvement in habitat availability — resulted in increased population sizes and subsequent accumulation of genetic diversity. This indicates that severe population size reductions have been absent, in spite of drastic short-term, high-amplitude climatic oscillations that appear to have occurred during late Pleistocene (see Roy *et al.*, 1996).

A closer look at the haplotype network (Fig. 2a) reveals some differences in the branching patterns of clades A and B; while essentially two groups of haplotypes exist in clade A (common and rare), in clade B haplotypes are more evenly distributed (Table 2) and, clade A displays a branching pattern in which haplotypes are separated by mostly one mutational steps. The more interior haplotypes (18, 19 and 29) are also the ones that are most abundant and have the largest geographical ranges. Other haplotypes seem to have derived from these, occur in low frequencies and have restricted geographical ranges. On the other hand, clade B shows a more linear and scattered branching pattern with several missing nodes. The most central haplotype (14) is not the most abundant and has a scattered geographical distribution. Among the more frequent haplotypes such as haplotypes 2 and 4, large as well as restricted geographical ranges can be found. According to Neigel & Avise (1993) lineage age and lineage range are correlated with older lineages having broader ranges. Therefore, our observations suggest a history of large and stable populations associated with clade A, allowing the diversification and maintenance of many haplotype variants. A history of small or fluctuating and possibly fragmented populations is more in line with the phylogeographical pattern observed for clade B.

# Historical biogeography

The first evidence for rabbit mtDNA geographical subdivision triggered the suggestion of a glacial refugium in the Mediterranean coast of France (Biju-Duval et al., 1991). More recently, the first northern Iberian populations were surveyed revealing an extensive variation inside clade B thus prompting Monnerot et al. (1994) to suggest that an isolate somewhere in the northern Iberian Peninsula was more plausible than one in southern France. The study of ancient mtDNA revealed that until the Middle Ages, when the natural range extended up to river Loire (Callou, 1995), French populations were characterized by haplotype 2 (clade B). During the Middle Ages rabbits quickly expanded all over France and with them mitochondrial type 1 (also clade B) (Hardy et al., 1995). Present day distribution of mtDNA types confirms the generalized occurrence of haplotypes 1 and 2, being haplotype 1 the most abundant (Mougel, 1997; Monnerot, 1998). Additionally, French populations are characterized by low levels of genetic variability at protein and immunoglobulin loci (van der Loo et al., 1991; Ferrand, 1995; Branco et al., 1999; van der Loo et al., 1999).

The data on genetic differentiation and diversity now available leads us to postulate that the glacial refugia for the rabbit were on the Iberian Peninsula one in the southwest and another one in the Mediterranean coast that probably extended to the north-east. Quaternary glaciations in the Iberian Peninsula appear to have been restricted to the Cordillera Cantábrica and Cordillera Central, leaving the southern region and the Mediterranean coastal area free of ice. A refugial area on the eastern coast of the Iberian Peninsula has already been suggested in phylogeographical studies of the European grasshoper (Chorthippus parallelus) (Cooper et al., 1995) and the Mediterranean ragwort (Senecio gallicus) (Comes & Abbot, 1998). The valley of the river Ebro may have formed a northward extension of the Mediterranean coast refuge. However, due to the proximity of the main glaciation areas of Cordillera Cantábrica, Cordillera Central and Pyrenees, the populations in the north-east refugium may have been subjected to more extreme climatic conditions than those in the south-west, possibly explaining the observed trend for high vs. low levels of genetic structuring that is observed in Fig. 2(b).

The observed phylogeographical pattern is that of two groups of populations with contiguous geographical ranges which overlap to a certain extent. The area of range overlap is a relatively narrow region crossing the Iberian Peninsula in a south-east-north-west direction. We observed a trend for haplotypes interior to the clades to be found at or near the proposed glacial refugia, while haplotypes with exterior positions were more frequently found in the region of overlap. Because a relative temporal scale can be inferred from the hierarchy of haplotype position in a phylogenetic tree whereby most interior haplotypes are the oldest and those with outer positions are younger (Templeton et al., 1995), the described pattern suggests that the south-east-north-west area across the Iberian Peninsula was colonized more recently giving rise to a secondary contact zone between rabbits carrying clades A and B (Fig. 1). This hypothesis is supported by the analysis of DNA extracted from rabbit remains dated from 11 500 to 4500 B.P., as only clade B haplotypes were detected in Central Iberian Peninsula and in the southern foothills of Sierra Nevada (Loreille et al., 1997; Monnerot, unpublished results). Thus, the contact in these areas probably does not exceed 4500 years.

Some clade B haplotypes were observed in the southwest of the Iberian Peninsula but only the most common

and widespread B haplotype (2) was detected. This observation may indicate a rapid penetration of rabbits approximately along the route Badajoz – Vila Viçosa – Infantado (Fig. 1). Conversely, a few A haplotypes were found in the north. Their local presence may have natural causes, but introductions by humans are highly probable in this region. Despite a low success rate (Branco et al., 1997), rabbit translocations for hunting purposes are known to have occurred in this region, mostly with animals from the southern regions where they are more abundant.

#### Intraspecific taxonomy

The data on cytoplasmic and nuclear genes (van der Loo et al., 1991; Ferrand, 1995; Branco & Ferrand, 1998; van der Loo et al., 1999) reveal the presence of two distinct and highly diversified evolutionary lineages of the European rabbit and support the recognition of two subspecies in the modern context of an appreciable level of genetic differentiation and historical continuity of barriers to genetic exchange (see Smith et al., 1997; Templeton, 1999). The A and B clades can then be correlated with, respectively, O. c. algirus and O. c. cuniculus. However, the match with traditional taxonomy is not perfect. For example only O. c. algirus was considered present on the Iberian Peninsula as morphological criteria were utilized for the description of rabbit subspecies (Cabrera, 1914). Thus, any future attempts to re-evaluate the taxonomy of O. cuniculus should take the genetic data into account.

## **Conclusions**

We observed (i) a strong discontinuity in the geographical distribution of two distinct clades A and B, that follows a north-west to south-east axis across the Iberian Peninsula (ii) a relatively narrow region along this axis where both clades are present in similar frequencies, and (iii) high levels of within-clade genetic diversity and structuring. We interpret the observed phylogeographical pattern as resulting from a secondary contact of two groups of rabbits after a long-term separation during which a considerable level of diversity accumulated between and within groups. This scenario is compatible with the occurrence of range contractions to two glacial refugia followed by range expansions in response to Quaternary climatic oscillations, resulting in the present day limited range overlap and admixture.

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